

ROLE OF REDOX AND CERULOPLASMIN IN IRON DEPOSITION IN GLIAL CELLS: IMPLICATION IN NEURODEGENERATIVE DAMAGES

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Received on: 05th-Mar-2011; Revised on: 14th-Mar-2010; Accepted on: 16th-Mr-2010; Published on: 3rd –Jul-2011 *Corresponding author: Email: ckm2300@mail.jnu.ac.in, mukhopc@yahoo.com; Tel: 011-26738738; Fax: 011-26741781

ABSTRACT

The cellular oxidation and reduction (redox) environment is influenced in presence of transition metals mainly iron and copper. They are also part of the regimen responsible for production and removal of reactive oxygen species (ROS). Interestingly, in most of the neurodegenerative diseases increased ROS generation and iron deposition were detected. However, their intrinsic relations either to cause the pathogenic condition found in neurodegenerative diseases or they are produced as a result of the condition is not clear yet. The human brain comprises only 2% of the total body weight, yet it is especially prone to ROS generation as it consumes about 20% of the resting total body oxygen. Similarly, need of glucose is also higher in active brain. Both the oxygen metabolism and glucose metabolism to gain energy are highly dependent on cellular iron metabolism. However, brain iron metabolism is so far less understood compare to the other organs. Since, ROS in presence of excess iron is highly reactive to cause oxidative damage, expression of iron homeostasis genes are usually regulated to avoid their proximity to each other. Glial cells play important role in movements of nutrients including essential metals like iron and copper to neurons as well as controlling ROS generation. Thus, it is important to understand the iron homeostasis components of glial cells in order to understand the role of redox/ROS and iron/copper mediated neurodegeneration. Ceruloplasmin (Cp) as a multicopper protein having ferroxidase (Fe₂+ to Fe₃+) activity performs a central role in body iron homeostasis. It has been described both as an antioxidant and oxidant molecule. In mammals, astroglia contains specialized membrane bound glycosyl-phosphatidyl-inositol (GPI)- anchored form of Cp that plays an important role in iron metabolism in central nervous system (CNS) by regulating iron release by maintaining stability of ferroportin. Mutation in Cp leads to iron deposition in various regions of CNS. All these evidences show a crucial role of Cp in maintaining body iron homeostasis including CNS. Here, we discuss the regulation of GPI-Cp by ROS that may be one of the potential mechanisms of iron deposition in glial cells.

Key words: Endothelial dysfunction; reactive oxygen species; oxidative stress; anti oxidants; nitric oxide; drug toxicity

[I] REDOX, METALS AND NEURO-DEGENERATION

Red-ox reactions represent the transfer of electrons from an electron donor (reducing agent) to an electron acceptor (oxidizing agent). The cellular redox environment is a balance between the production of reactive oxygen species (ROS) and their removal by antioxidant enzymes and small molecular weight antioxidants. ROS are oxygen-containing molecules that are highly reactive. The partial reduction of molecular oxygen results in the production of superoxide (O2.-) and hydrogen peroxide (H₂O₂) [1]. O₂.- and H₂O₂ react with transition metal ions (e.g., iron and copper) through Fenton and Haber-Weiss chemistry, generating the highly reactive hydroxyl radical (HO.) [2]. Redox-active metals catalyze many essential reactions for brain function as cofactors for specific enzymes, participate in

electron transfer reactions required for cellular metabolism and oxygen transport [3, 4]. However, these metals can also participate in the generation of highly toxic free radicals that can cause oxidative damage to cells [5]. Iron-induced oxidative stress is par-ticularly dangerous because it can cause further iron release from iron-containing proteins, such as ferritin (Ft), heme proteins and iron-sulfur (Fe-S) clusters, forming a positive-feedback destructive intracel¬lular loop that exacerbates the toxic effects of brain iron overload [6]. Brain iron overload is cause of, or has been associated with the development of several neurodegenerative diseases including Parkinson's, Friedreich's ataxia, aceruloplasminemia, pantothenate kinase deficiency and others. Neurodegenerative diseases and their association in iron deposition reported in the literature have summarized below in Table-1.

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Table: 1. Neurodegenerative diseases and their association in iron deposition.

No	Neurodegenerative disease	Refs
1	Alzheimer's disease (AD)	[7-9]
2	Parkinson's disease (PD)	[8, 10, 11]
3	Multiple sclerosis (MS)	[12, 13]
4	Friedreich's ataxia	[14, 15}
5	Huntington's disease	[16, 17]
6	Aceruloplasminemia	[18]
7	Amyotrophic lateral sclerosis (ALS)	[19-21]
8	Hallervorden-Spatz syndrome (HSS)	[22, 23]
9	Neuroferritinopathy	[24, 25]

SPECIES REACTIVE OXYGEN AND F 11 1 NEURODEGENERATION

ROS are byproducts of metabolism and considered as dangerous towards biological materials because of their roles in generating hydroxyl radical in conjunction with transition metals like iron and copper by Fenton's reaction. ROS are implicated in several metabolic disorders like atherosclerosis. cancer. neurodegenerative diseases, aging as well as in infectious diseases [26]. However, more recently, an essential role of ROS in many cellular signaling events was identified [27]. The net ROS generation is a balance between total generation and amount consumed by various non-enzymatic or enzymatic antioxidants. Various conditions leading to increased cellular ROS generation or depletion of antioxidants result into net increase in ROS generation.

Brain is thought to be particularly susceptible to ROS accumulation because of its high utilization of oxygen for metabolic processes and its relative paucity of antioxidant and regenerative properties compared to other organs [28]. ROS may arise from any number of normal or dysfunctional cellular mechanisms including auto-oxidation of catecholamines [29], disruption of mitochondrial complexes [30], inappropriate incorporation of exogenous toxins, inadequate availability of glutathione (GSH) or improperly stored or excess concentrations of free iron or copper by Fenton reaction [26, 28, 31]. Since, ROS can oxidize vital cellular components such as lipids, proteins and nucleic acid; it may cause cellular damage and subsequent cell death. The lack of regeneration capacity of neurons, once they are degenerated may lead to pathogenic conditions like AD, PD, ALS or others [32].

ROS also accumulates in brain due to exposure of pesticides (paraquat, diquat, maneb, rotenone, organochlorines) as suggested by epidemiological studies demonstrating a relationship between pesticide exposure and brain neurodegeneration [33]. Agricultural toxin paraquat is a potential neurotoxin as it has the ability to cross the blood brain barrier [34, 35]. Recent evidences show that in response to certain environmental toxins and endogenous proteins, microglia release ROS that cause neurotoxicity [36]. Strong evidences are provided to show that microglia identifies neurotoxic stimuli through pattern recognition receptors (PRRs) and activates NADPH oxidase activity to generate ROS [36]. In response to rotenone [37], paraquat [38, 39], lipopolysaccharide [40, 41], αsynuclein [42, 43], amyloid- β [44, 45], diesel exhaust particles [46] and others [36] microglia could be activated to generate NADPH oxidase induced ROS.

Glutathione is the major antioxidant present in brain tissue and the most important redox buffer in cells [47-49]. Glutathione is present in the brain in millimolar (mM) concentration [49]. Glutathione peroxidase (Gpx) is the major enzyme for the detoxification of H2O2 in the brain since the brain has comparatively lower catalase activity. Interestingly, GSH concentration appears to be higher in astrocytes than neurons [49]. Although varying in different regions of the brain, all GSH levels diminish by about 30% in age related diseases [50] suggesting a possible link with the increased ROS generation reported in AD and PD [Table-2]. Depletion of GSH may render cells more sensitive to toxic effects of oxidative stress and potentiate the toxic effects of reactive microglia [49, 51, 52]. Information on the origin of brain GSH and its possible transport from blood to brain is limited. A substantial uptake of 35S-labeled GSH by rat brain was found suggesting that GSH can cross blood brain barrier (BBB) by a saturable and specific mechanism [53]. Heme oxygenase-1 (HO-1) expression appears to be an excellent marker of oxidative stress related to cell injury in the brain [54] as GSH depletion induces HO-1 in the brain. Elevated GSH levels in hippocampus and midbrain were also reported in AD [55], an indication that AD neurons may be over-reacting to an oxidative load. Similarly, decreased activity of antioxidant enzymes occurs in AD brains [56], an indication that the normal handling of GSH may be altered in these cells. A 30-40% decrease in GSH concentrations without a corresponding increase in the levels of oxidized GSH (GSSG) was also reported in PD brains [57]. In almost all cases of neurodegenerative diseases substantial increase in net ROS levels were reported [Table-2].

Table: 2. ROS and neurodegenerative diseases.



	Neurodegenerative disease	
No		References
1	Alzheimer's disease (AD)	[58-63]
2	Parkinson's disease (PD)	[61, 64-66]
3	Amyotrophic lateral sclerosis (ALS)	[67-70]
4	Huntington's disease	[71-75]
5	Friedreich's ataxia	[76-79]
6	Multiple sclerosis (MS)	[80-83]
7	Aceruloplasminemia	[84-85]
8	Neuroferritinopathy	[86]

[III] CERULOPLASMIN

Ceruloplasmin (Cp), a copper containing 132-kDa acute phase α 2-glycoprotein regulates body iron homeostasis by its capacity as a ferroxidase [87-88]. It binds ~95% of copper found in human plasma and is mainly synthesized and secreted from the liver [89, 90]. In the central nervous system of humans and other mammals, Cp is expressed in astroglial cells as a GPI-anchored membrane bound form [91, 92]. Cp was first isolated from plasma and characterized as a copper containing protein by Holmberg and Laurell in 1948 [93]. Other than liver and brain organs expressing Cp gene are eyes, lungs, spleen and testis [94-96]. In 1984, Putnam determined the complete amino acid sequence of human ceruloplasmin, revealing the single-chain structure of this molecule [97]. As a major ferroxidase in plasma, Cp catalyzes conversion of Fe2+ to Fe3+ for binding to apo-transferrin [98]. The role of Cp in iron homeostasis is confirmed by findings of abnormal iron metabolism in patients with hereditary Cp deficiency [99] and in mice with targeted disruption of the Cp gene [100]. Patients with aceruloplasminemia have impaired iron export from certain tissues and characterized by iron overload in retina, brain and pancreas [85,101]. Also Cp-/- mice exhibit similar iron overload in brain and other visceral organs [100,102,103]. These findings, together with early organ culture studies [98, 104] suggest that Cp is required for efficient iron release from cells and tissues. In contrast, Cp has been shown to mediate inward iron flux as well in several cell culture systems including hepatic, erythroid [105,106] and glioblastoma cells [107,108]. Iron deposition in of aceruloplasminemia patients and related brain neurodegeneration strongly indicate its role as a neuroprotector in central nervous system by regulating iron transport. The ability of GPI-Cp in astrocytes to release iron was confirmed using purified astrocytes from Cp knock-out mice [109]. Subsequent studies to reveal the role of Cp in iron release illustrated that GPI-Cp co-localizes on the astrocyte cell surface with a ferrous iron transporter, ferroportin (IREG1). A recent study shows the ferroxidase activity of GPI-Cp is required for stability of ferroportin providing a molecular mechanism of iron deposition in brain in absence of or in reduced content of Cp [110]. Any reduction of Cp may, thus, affect cellular release of iron and cause oxidative damages in presence of ROS.

Besides its role in iron homeostasis, Cp is also reported to have other functions including participation in several biological oxidation reactions that include role in copper transport, coagulation, angiogenesis, defense against oxidant stress as antioxidant and role in low density lipoprotein oxidation [111]. Cp was described as an antioxidant because of its ability to inhibit the oxidation of lipids [26] as well as for its ability to scavenge superoxide radical (O2.-) and sequestering of free copper ions [112]. The ferroxidase activity may also contribute to the antioxidant capacity of Cp, because conversion of Fe2+ to Fe3+ may reduce oxidant capacity of iron by inhibition of the Fenton reaction. In contrast, several other studies have shown that Cp to contain pro-oxidant activity and ability to oxidize low density lipoprotein (LDL) in presence of vascular cells like endothelial, smooth muscle cells or monocytes and implicated in atherosclerosis [111, 113, 114]. Recently, its role as a nitrite oxidase has also been established [115].

3.1. Gene structure

Human Cp is encoded by 20 exons encompassing approximately 65 kb of DNA localized to chromosome 3q23-q24 [116, 117]. In hepatocytes, Cp gene is expressed as two transcripts of 3.7 and 4.2 kb, which arise from use of alternative polyadenylation sites within the 3' untranslated region [118]. Cloning and characterization of Cp from rat and mouse reveals 90% amino acid homology with the human sequence and similar patterns of gene expression in all three species [94, 95]. Within the human central nervous system Cp is expressed in astrocytic glia lining the brain microvasculature, surrounding dopaminergic neurons in the substantia nigra and within the inner nuclear layer of the retina [91]. Recent studies demonstrate that Cp is synthesized as a GPI-anchored protein generated by alternative splicing of exons 19 and 20 in astrocytes, sertoli cells and lymphocyte [92, 119-121, 122]. As a result, the 5 C-terminal amino acids found in secretory form of Cp are replaced by a 30-amino acid stretch in GPI-Cp. The spatial structure of human Cp and the precise total amount of six copper ions in its molecule were elucidated when a crystallographic picture at 3.1A° resolution was obtained [123]. Although copper has no effect on the rate of synthesis or secretion of Cp, failure to incorporate this metal during synthesis results in the secretion of an unstable apoceruloplasmin moiety that is devoid of ferroxidase activity [124, 125].

3



3.2. Ceruloplasmin and neurodegeneration

Cp is a key protein involved in the regulation of the redox state of iron by converting the ROS catalytic Fe(II) to a less reactive Fe(III) by virtue of its ferroxidase activity. Iron deposition in brain aceruloplasminemia patients and of related neurodegeneration strongly indicate its role as a neuroprotector in the central nervous system by regulating iron transport [126-130]. Initially, it was suggested that GPI-anchored Cp in astrocytes could promote iron release [102] that was later confirmed using purified astrocytes from Cp knock-out mice [109]. Ferroportin is a unique and ubiquitous iron exporter of mammalian cells including astroglia [131]. GPI-anchored Cp is co-localized on membrane of astrocytes with ferroportin. It was demonstrated that ferroxidase activity of Cp is required for the stability of ferroportin [110]. Cp-knockout mice exhibit severe defects in iron release from astrocytes, probably resulting from the lack of ferroxidase activity, which is necessary for the exporter function and stability of ferroportin. In absence of Cp, ferroportin loses its ability to export iron that may explain iron accumulation in astroglia in aceruloplasminemia. Taken together, these results suggest a role for Cp in the regulation of cellular iron efflux implying its role in the pathogenesis of neurodegeneration involving increased iron and oxidative damage, such as PD and AD.

3.3. Reactive oxygen species decrease ceruloplasmin expression

We recently demonstrated a novel negative regulation of Cp synthesis by ROS in rat C6 glial and human astroglia U373MG cells by mRNA decay mechanism. Cp is reported to predominantly express a GPI-anchored membrane bound form in glial cells [103]. We demonstrated that ROS generated either intracellularly by inhibition of mitochondrial electron transport chain as may happen by environmental toxins or extracellularly as may be generated by NADPH oxidases of activated macrophages, neutrophils or microglial cells could decrease Cp synthesis. The study further revealed the involvement of its 3'untranslated region (3'UTR) in ROS mediated regulation of Cp as verified by conferring a promotion of mRNA decay using heterologous reporter, where addition of Cp 3'UTR downstream of CAT gene cause decay of CAT mRNA in astroglial cells [130]. We further demonstrated that in response to ROS, a decrease in binding of yet unidentified protein to 3'UTR makes it apparently susceptible to endonuclease mediated cleavage. The complete blocking of the reduction of RNA-protein complex by antioxidant N- acetyl cysteine shows the actual role of ROS is to regulate the complex formation of the protein with the Cp 3'UTR.

Increase in cellular ROS generation was previously shown to increase HO-1 content suggesting HO-1 mediated heme degradation during ROS generation [132]. The resultant increase in intracellular labile iron pool (LIP) was confirmed by EPR analysis [132]. Increase in cellular ferritin synthesis is reported in hepatic cells probably to protect cells from iron-mediated cellular damage by storing the excess intracellular iron [132]. In

contrast, both the ferritin-H and-L chains are degraded in presence of ROS in microglial cells [133]. In fact, that would also increase the intracellular iron pool and may lead to iron mediated injury. To avoid this iron-mediated injury Cp should help release iron through ferroportin. There is so far no report on ferroportin status in astrocytes, microglia or neurons by ROS. In fact, our work shows GPI-Cp is decreased in presence of ROS that would affect ferroportin status and resultant increase in intracellular iron pool.

The presence of AU-rich responsive element (ARE) or stemloop structure like iron responsive element (IRE) is often reported in 3'UTR of genes those are regulated by mRNA decay/stability mechanism [134]. The absence of any ARE or IRE in Cp 3'UTR opens the intriguing possibility of finding a novel response element involving mRNA stability/decay mechanism in mammalian cells. We hypothesize that a redox protein normally remains bound to the 3'UTR and provides stability to Cp transcript in glial cells. In response to ROS, this redox-sensitive protein may undergo oxidative modification and eventually leaves 3'UTR. As a result, the unoccupied 3'UTR becomes a better substrate for endonuclease cleavage. The region of Cp 3'UTR responsible for binding the protein and the mechanism by which ROS affect the binding of this protein remains to be determined.

This ROS mediated regulation of Cp could also explain iron accumulation and related injury in neurodegenerative diseases. Generations of ROS in neuronal and glial cells by inflammation, injury or by environmental toxins like pesticides [Table-2] are implicated in developing these neurodegenerative diseases [36, 135-137]. Thus, in a condition, when ROS generation is increased either by environmental toxins like pesticides or by inflammation, concomitant decrease in GPI-Cp synthesis in glial would result into accumulation of iron within the cell cells probably by simultaneous decrease of ferrous iron transporter ferroportin as described recently [110]. Thus, generated ROS and resultant accumulated iron can form highly reactive hydroxyl radical by Fenton reaction and damage glial cells. The role of glial cells is well appreciated in neuroprotection [138]. Therefore, any damage in glial cells can lead into damages of associated neurons. Thus, our finding could explain how neuronal damage might happen by increased ROS generation in glial cells by environmental toxins or other pathological conditions, a likely scenario in most of the neurodegenerative diseases. Very recent demonstration of increased neurotoxicity and lipid peroxide products in brains of rotenone treated, Cpdeficient mice strongly support our finding [139].

[IV] CONCLUSION AND FUTURE PERSPECTIVE

Iron plays a crucial role in maintaining several functions of brain but in increasing concentration can act as a catalyst for detrimental oxidative damages to elevate the chances of neurodegenerative diseases. Generation of ROS and iron deposition both are found to be increased in most of the edited BY – prof. S.K Maulik; MD, phd



neurodegenerative diseases. Although role of iron in elevating oxidative damage in conjunction with ROS is well appreciated, the role of ROS in dysregulation of iron homeostasis has not been explored much. The recent finding of down-regulation of GPI-Cp by ROS thus opened a new avenue of understanding iron deposition detected in neurodegenerative diseases. The detail molecular mechanism of this novel mRNA decay mechanism may identify newer players important for maintaining iron homeostasis in brain. The new knowledge and technology of proteomics and bioinformatics will be highly helpful to identify these new molecular players responsible for iron deposition in brain. This knowledge also may be helpful in predicting iron deposition in individuals by examining status of these molecules in younger age of any individual. Developing newer iron chelators that can cross blood brain barrier may also open novel therapeutic strategies to prevent or slow the progression of these neurodegenerative diseases.

REFERENCES

- [1] Halliwell B, Gutteridge JMC. [1999] Free radicals in biology and medicine. 3rd ed. New York: *Oxford University Press* p:936.
- [2] Halliwell B, Gutteridge JMC. [1992] Biologically relevant metal ion-dependent hydroxyl radical generation: an update. *FEBS Lett* 307: 108–112.
- [3] Andrews NC, Schmidt PJ. [2007] Iron homeostasis. *Annu Rev Physiol* 69: 69–85.
- [4] Balamurugan K, Schaffner W. [2006] Copper homeostasis in eukaryotes: teetering on a tightrope. *Biochim Biophys Acta* 1763: 737-46.
- [5] Perez CA, Tong Y, Guo M. [2008] Iron chelators as potential therapeutic agents for Parkinson's disease. *Curr Bioact Compd* 4: 150–158.
- [6] MacKenzie EL, Iwasaki K, Tsuji Y. [2008] Intracellular iron transport and storage: from molecular mechanisms to health implications. Antioxid. *Redox Signal* 10: 997–1030.
- [7] Connor JR, Snyder BS, Beard JL, Fine RE, Mufson EJ. [1992] Regional distribution of iron and iron-regulatory proteins in the brain in aging and Alzheimer's disease. J Neurosci Res 31: 327–335.
- [8] Qian ZM, Wang Q. [1998] Expression of iron transport proteins and excessive iron accumulation in the brain in neurodegenerative disorders. *Brain Res Rev* 27: 257–67.
- [9] Thompson K, Menzies S, Muckenthaler M, et al. [2003] Mouse brains deficient in H-ferritin have normal iron concentration but a profile of iron deficiency and increased evidence of oxidative stress. *J Neurosci Res* 71: 46–63.
- [10] Dexter DT, Wells FR, Agid F, Agid Y, Lees AJ, Jenner P, et al. [1987] Increased nigral iron content in postmortem parkinsonian brain. *Lancet* 2: 1219–20.
- [11] Riederer P, Sofic E, Rausch W, et al. [1989] Transition metals, ferritin, glutathione, and ascorbic acid in parkinsonian brains. *J Neurochem* 52: 515–520.
- [12] LeVine SM. [1991] Oligodendrocytes and myelin sheaths in normal, quaking and shiverer brains are enriched in iron. J Neurosci Res 29: 413–419.
- [13] Bakshi R, Dmochowski J, Shaikh ZA, Jacobs L. [2001] Gray matter T2 hypointensity is related to plaques and atrophy in

the brains of multiple sclerosis patients. J Neurol Sci 185: 19–26.

- [14] Connor JR, ed. [1997] Metals and oxidative damage in neurological disorders. *New York: Plenum Press.*
- [15] Waldvogel D, van Gelderen P, Hallett M. [1999] Increased iron in the dentate nucleus of patients with Friedrich's ataxia. *Ann Neurol* 46: 123–125.
- [16] Chen JC, Hurdy DA, Hucharczyk W, et al. [1993] MRI of human postmortem brain tissues correlative study between T2 and assays of iron and ferritin in Parkinson's and Huntington's disease. *American Journal of Neurological Research* 14: 275–281.
- [17] Bartzokis G. Magnetic resonance imaging of brain iron. In: Connor JR. (Ed.). [1997] Metals and Oxidative Damage in Neurological Disorders. *Plenum Press, New York.* pp. 41–56.
- [18] Miyajima H. [2003] Aceruloplasminemia, an iron metabolic disorder. *Neuropathology* 23: 345–350.
- [19] Oba H, Araki T, Ohtomo K, et al. [1993] Amyotrophic lateral sclerosis: T2 shortening in motor cortex at MR imaging. *Radiology* 189: 843–846.
- [20] Kasarskis EJ, Tandon L, Lovell MA and Ehmann WD. [1995] Aluminum, calcium, and iron in the spinal cord of patients with sporadic amyotrophic lateral sclerosis using laser microprobe mass spectroscopy: a preliminary study J *Neuro Sci* 130: 203–208.
- [21] Carri MT, Ferri A, Cozzolino M, et al. [2003] Neurodegeneration in amyotrophic lateral sclerosis: the role of oxidative stress and altered homeostasis of metals. *Brain Res Bull* 61: 365–374.
- [22] Ponting CP. [2001] Domain homologues of dopamine βhydroxylase and ferric reductase: roles for iron metabolism in neurodegenerative disorders? *Hum Mol Genet* 10: 1853–58.
- [23] Zhou B, Westaway SK, Levinson B, et al. [2001] A novel pantothenate kinase gene (PANK2) is defective in Hallervorden-Spatz syndrome. *Nat Genet* 28: 345–349.
- [24] Curtis AR, Fey C, Morris CM, et al. [2001] Mutation in the gene encoding ferritin light polypeptide causes dominant adult-onset basal ganglia disease. *Nat Genet* 28: 350–354.
- [25] Ponka P. [2002] Rare causes of hereditary iron overload. *Semin Hematol* 39: 249–262.
- [26] Halliwell B, Gutteridge JMC. [1990] The antioxidants of human extracellular fluids. Arch Biochem Biophys 280: 1–8.
- [27] Son Y, Cheong YK, Kim NH, Chung HT, Dae Kang G, Pae HO. [2011] Mitogen-activated protein kinases and reactive oxygen species: How can ROS activate MAPK pathways? *Journal of Signal Transduction* 2011:1–6.
- [28] Rhodes SL, Ritz B. [2008] Genetics of iron regulation and the possible role of iron in Parkinson's disease. *Neurobiology of Disease* 32: 183–19.
- [29] Behonick GS, Novak MJ, Nealley EW, Baskin SI. [2001] Toxicology update: the cardiotoxicity of the oxidative stress metabolites of catecholamines (aminochromes). *J Appl Toxicol* 21: S15–22.
- [30] Chandel NS, McClintock DS, Feliciano CE, et al. [2000] Reactive oxygen spe-cies generated at mitochondrial complex III stabilize hypoxia-inducible factor-1α during hypoxia: a mechanism of O2 sensing. *J Biol Chem* 275: 25130–25138.
- [31] Bharath S, Hsu M, Kaur D, Rajagopalan S, Andersen JK.[2002] Glutathione, iron and Parkinson's disease. Biochemical Pharmacology 64: 1037–1048.
- [32] Andersen JK. [2004] Oxidative stress in neurodegeneration: Cause or consequence? *Nat Med 10(Suppl.):* S18–S25.

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- [33] Francoa R, Sumin Li, Rodriguez-Rocha H, Burns M, Panayiotidis MI, et al. [2010] Molecular mechanisms of pesticide-induced neurotoxicity: Relevance to Parkinson's disease. *Chemico-Biological Interactions* 188: 289–300.
- [34] Corasaniti MT, Strongoli MC, Pisanelli A, et al. [1992] Distribution of paraquat into the brain after its systemic injection in rats. *Funct Neurol* 7: 51–56.
- [35] Widdowson PS, Farnworth MJ, Simpson MG, Lock EA. [1996] Influence of age on the passage of paraquat through the blood-brain barrier in rats: a distribution and pathological examination. *Hum Exp Toxicol* 15: 231–236.
- [36] Block ML, Zecca L, Hong JS. [2007] Microglia-mediated neurotoxicity: uncovering the molecular mechanisms. *Nature reviews/ neuroscience* 8: 57–69.
- [37] Gao HM, Hong JS, Zhang W, Liu B. [2002] Distinct role for microglia in rotenone-induced degeneration of dopaminergic neurons. J Neurosci 22: 782–790.
- [38] Wu XF, Block ML, Zhang W, Qin L, Wilson B, Zhang WQ et al. [2005] The role of microglia in paraquat- induced dopaminergic neurotoxicity. *Antioxid Redox Signal* 7: 654– 661.
- [39] Bonneh-Barkay D, Reaney SH, Langston WJ, Di Monte DA. [2005] Redox cycling of the herbicide paraquat in microglial cultures. *Brain Res Mol Brain Res* 134: 52–56.
- [40] Gao H, Jiang J, Wilson B, et al. [2002] Microglial activationmediated delayed and progressive degeneration of rat nigral dopaminergic neurons: relevance to Parkinson's disease. J Neurochem 81: 1285–1297.
- [41] Qin L, Liu Y, Wang T, et al. [2004] NADPH oxidase mediates lipopolysaccharide-induced neurotoxicity and proinflammatory gene expression in activated microglia. J Biol Chem 279: 1415–1421.
- [42] Zhang W, Wang T, Pei Z, et al. [2005]. Aggregated α -synuclein activates microglia: a process leading to disease progression in Parkinson's disease. *FASEB J* 19: 533–542.
- [43] Croisier E, Graeber MB. [2006] Glial degeneration and reactive gliosis in α -synucleinopathies: the emerging concept of primary gliodegeneration. *Acta Neuropathol (Berl)* 112: 517–530.
- [44] Wilkinson B, Koenigsknecht-Talboo J, Grommes C, Lee CY, Landreth G. [2006] Fibrillar β-amyloid-stimulated intracellular signaling cascades require Vav for induction of respiratory burst and phagocytosis in monocytes and microglia. J Biol Chem 281: 20842–20850.
- [45] Qin L, Liu Y, Cooper C, Liu B, Wilson B, Hong JS. [2002] Microglia enhance β-amyloid peptide-induced toxicity in cortical and mesencephalic neurons by producing reactive oxygen species. *J Neurochem* 83: 973–983.
- [46] Block ML, Wu X, Pei Z, Li G, Wang T, Qin L, et al. [2004] Nanometer size diesel exhaust particles are selectively toxic to dopaminergic neurons: the role of microglia, phagocytosis and NADPH oxidase. *FASEB J* 18: 1618–1620.
- [47] Meister A, Andersson M. [1983] Glutathione. Annu Rev Biochem 52: 711–760.
- [48] Reynolds A, Laurie C, Mosley RL and Gendelman HE. [2007] Oxidative stress and the pathogenesis of neurodegenerative disorders. *International review of neurobiology* 82: 297–325.
- [49] Dringen R, Gutterer JM and Hirrlinger J. [2000] Glutathione metabolism in brain metabolic interaction between astrocytes and neurons in the defense against reactive oxygen species. *Eur J Biochem* 267: 4912–4916.

- [50] Chen TS, Richie JP, Lang CA. [1989] The effect of aging on glutathione and cysteine levels in different regions of the mouse brain. *Proc Soc Exp Biol Med* 190: 399–402.
- [51] Sian J, Dexter DT, Lees AJ, et al. [1994] Alterations in glutathione levels in Parkinson's disease and other neurodegenerative disorders affecting basal ganglia. *Ann Neurol* 36: 348–355.
- [52] Winterbourn CC, Metodiewa D. [1994] The reaction of superoxide with reduced glutathione. *Arch Biochem Biophys* 314: 284–290.
- [53] Kannan R, Uuhlenhamp JF, Jaendidier E, Trinh H, Oouhbtens M, Laplowitz N. [1990] Evidence for carriermediated transport of glutathione across the blood brain barrier in the rat. *Journal of Clinical Investigation* 85: 2009– 2013.
- [54] Sharp FR. [1995] Stress proteins are sensitive indicators of injury in the brain produced by ischemia and toxins. *J Toxicol Sci* 20: 450–453.
- [55] Adams JD Jr, Klaidmen LK, Odunze IN, Shen HC, Miller CA. [1991] Brain levels of glutathione, glutathione disulfide, and vitamin E. *Mol Chem Neuropathol* 14: 213–226.
- [56] Omar RA, Chyan YJ, Andorn AC, Poeggler B, Robakis NK, Pappola MA. [1999] Increased expression but reduced activity of antioxidant enzymes in Alzheimer's Disease. J Alzheimer's Dis 1: 39–145.
- [57] Perry TL, Yong VW. [1986] Idiopathic Parkinson's disease, progressive supranuclear palsy and glutathione metabolism in the substantia nigra of patients. *Neurosci Lett* 67: 269–274.
- [58] Frautschy SA, Baired A, Cole GM. [1991] Effects of injected Alzheimer's β-amyloid cores in rat brain. *Proc Natl Acad Sci* USA 88: 8362–8366.
- [59] Markesbery WR. [1997] Oxidative stress hypothesis in Alzheimer's disease. *Free Radical Biology and Medicine* 23:134–147.
- [60] Multhaup G, Schlicksupp A, Hesse L, Beher D, Masters CL, Beyreuther K. [1997] Reactive oxygen species and Alzheimer's disease. *Biochem Pharmacol* 54: 533–539.
- [61] Tabner BJ, Turnbull S, Omar MA. El-Agnaf, Allsop D. [2003] Direct production of reactive oxygen species from aggregating proteins and peptides implicated in the pathogenesis of neurodegenerative diseases. *Curr Med Chem* - *Immun, Endoc & Metab Agents* 3: 299–308.
- [62] Reddy PH. [2006] Amyloid precursor protein- mediated free radicals and oxidative damage: Implications for the development and progression of Alzheimer's disease. *Journal of Neurochemistry* 96: 1–13.
- [63] Dumont M, Beal MF. [2010] Neuroprotective strategies involving ROS in Alzheimer's disease. *Free Radic Biol Med* 1–13.
- [64] Fahn S, Cohen G. [1992] The oxidant stress hypothesis in Parkinson's disease. Evidence supporting it. Annals of Neurology 32: 804–812.
- [65] Akaneya Y, Takahasi M, Hatanaka H. [1995] Involvement of free radicals in MPP+ neurotoxicity against rat dopaminergic neuron in culture. *Neuroscience Letters* 193: 53–56.
- [66] Cassarino DS, Fall CP, Swerdlow RH, et al. [1997] Elevated reactive oxygen species and antioxidant enzyme activities in animal and cellular models of Parkinson's disease. *Biochim Biophys Acta* 1362:77–86.
- [67] Rosen DR, Siddique T, Patterson D, et al. [1993] Mutations in Cu/Zn SOD gene are associated with familial amyotrophic lateral sclerosis. *Nature* 362: 59–62.

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[68] Robberecht W, Sapp P, Viaene MU, et al. [1994] Cu/Zn super-oxide dismutase activity in familial and soporadic amytrophic lateral sclerosis. *Journal of Neurochemistry* 62: 1384–1387.

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- [69] Liu D, Wen J, Liu J, Li L. [1999] The roles of free radicals in amyotrophic lateral sclerosis: reactive oxygen species and elevated oxidation of protein, DNA, and membrane phospholipids. *FASEB J* 13: 2318–2328.
- [70] Ahmed MS, Hung WY, Zu JS, Hockberger P, Siddique T. [2000] Increased reactive oxygen species in familial amyotrophic lateral sclerosis with mutations in SOD1. *Journal of the Neurological Sciences* 176: 88–94.
- [71] Beal MF. [1996] Mitochondria, free radicals and neurodegeneration. *Current Opinion in Neurobiology* 6: 661– 666.
- [72] Gu M, Gash MT, Mann VM, Jany-Agid F, Cooper JM, Schapira AH. [1996] Mitochondrial defect in Huntington's disease caudate nucleus. *Annals of Neurology* 39: 385–389.
- [73] Loeffler DA, Lewitt PA, Juneau PL, et al. [1996] Increased regional brain concentration of ceruloplasmin in neurodegenerative disorders. *Brain Research* 738: 265–274.
- [74] Browne SE, Bowling AC, MacGarrey U, et al. [1997] Oxidative damage and metabolic dysfunction in Huntington's disease: selective vulnerability of the basal ganglia. *Annals of Neurology* 41: 646–653.
- [75] Wyttenbach A, Sauvageot O, Carmichael J, et al. [2002] Heat shock protein 27 prevents cellular polyglutamine toxicity and suppresses the increase of reactive oxygen species caused by huntingtin. *Mol Genet* 11: 1137–1151.
- [76] Rotig A, de Lonlay P, Chretien D, et al. [1997] Aconitase and mitochondrial iron–sulphur protein deficiency in Friedreich ataxia. *Nat Genet* 17: 215–217.
- [77] Tozzi G, Nuccetelli M, Lo Bello M, et al. [2002] Antioxidant enzymes in blood of patients with Friedreich's ataxia. Arch Dis Child 86: 376–380.
- [78] Llorens JV, Navarro JA, Martinez SMJ, et al. [2007] Causative role of oxidative stress in a Drosophila model of Friedreich ataxia. *FASEB J* 21: 333–344.
- [79] Armstrong JS, Khdour O, Hecht SM. [2010] Does oxidative stress contribute to the pathology of Friedreich's ataxia? A radical question. *FASEB J* 24: 2152–2163.
- [80] Gilgun-Sherki Y, Melamed E, Offen D. [2004] The role of oxidative stress in the pathogenesis of multiple sclerosis: the need for effective antioxidant therapy. *J Neurol* 251: 261–8.
- [81] Mattson MB, Taub DD. [2004] Ancient viral protein enrages astrocytes in multiple sclerosis. *Nature neuroscience* 7: 1021–1023.
- [82] Miller E, Mrowicka M, Zolynski K, Kedziora J. [2009] Oxidative stress in multiple sclerosis. *Pol Merkur Lekarski* 27: 499–502.
- [83] Horssen JV, Witte ME, Schreibelt G and de Vries HE. [2011] Molecular Basis of Multiple Sclerosis. Biochimica et Biophysica Acta (BBA) - *Molecular Basis of Disease* 1812: 141–150.
- [84] Yoshida K, Kaneko K, Miyajima H, Tokuda T, Nakamura A, Kato M, Ikeda S. [2000] Increased lipid peroxidation in the brains of aceruloplasminemia patients. *Journal of the Neurological Sciences* 175: 91–95.
- [85] Miyajima H, Takahashi Y, Kono S. [2003] Aceruloplasminemia, an inherited disorder of iron metabolism. *Biometals* 16: 205–13.

- [86] Cozzi A, Rovelli E, Frizzale G, et al. [2010] Oxidative stress and cell death in cells expressing L-ferritin variants causing neuroferritinopathy. *Neurobiol Dis* 37: 77–85.
- [87] Osaki, S. [1966] Kinetic studies of ferrous ion oxidation with crystalline human ferroxidase (ceruloplasmin). J Biol Chem 241: 5053–5059.
- [88] Vassiliev V, Harris ZL, Zatta P. [2005] Ceruloplasmin in neurodegenerative diseases. *Brain Research Reviews* 49: 633–640.
- [89] Sato M, Gitlin JD. [1991] Mechanisms of copper incorporation during the biosynthesis of human ceruloplasmin. *J Biol Chem* 266:5128–34.
- [90] Lee DW, Andersen JK, Kaur D. [2006] Iron dysregulation and neurodegeneration: The molecular connection. *Molecular Interventions* 6: 89–97.
- [91] Klomp LW, Gitlin JD. [1996] Expression of the ceruloplasmin gene in the human retina and brain: implications for a pathogenic model in aceruloplasminemia. *Hum Mol Genet* 5: 1989–1996.
- [92] Patel BN, David S. [1997] A novel glycosylphosphatidylinositol- anchored form of ceruloplasmin is expressed by mammalian astrocytes. *J Biol Chem* 272: 20185–90.
- [93] Holmberg CG, Laurell CB. [1948] Investigations in serum copper. II. Isolation of the copper-containing protein and a description of some of its properties. *Acta Chem Scand* 2: 550–56.
- [94] Aldred AR, Grimes A, Schreiber G, Mercer JF. [1987] Rat ceruloplasmin. Molecular cloning and gene expression in choroids plexus, yolk sac, placenta, and testis. *J Biol Chem* 262: 2875–2878.
- [95] Fleming R, Gitlin JD. [1990] Primary structure of rat ceruloplasmin and analysis of tissue-specific gene expression during development. *J Biol Chem* 265: 7701–7707.
- [96] Klomp LWJ, Farhangrazi ZS, Dugan LL, Gitlin JD. [1996] Ceruloplasmin gene expression in the murine central nervous system. *J Clin Invest* 98: 207–215.
- [97] Takahashi N, Ortel TL, Putnam FW. [1984] Single-chain structure of human ceruloplasmin: the complete amino acid sequence of the whole molecule. *Proc Natl Acad Sci* USA 81: 390–94.
- [98] Osaki S, Johnson DA. [1969] Mobilization of liver iron by ferroxidase (ceruloplasmin). *J Biol Chem* 244: 5757–5765.
- [99] Miyajima H, Nishimura Y, Mizuguchi K, Sakamoto M, Shimizu T, Honda N. [1987] Familial apoceruloplasmin deficiency associated with blepharospasm and retinal degeneration. *Neurology* 37: 761–767.
- [100] Harris ZL, Durley AP, Man TK, Gitlin JD. [1999] Targeted gene disruption reveals an essential role for ceruloplasmin in cellular iron efflux. *Proc Natl Acad Sci USA* 96: 10812–7.
- [101] Yamaguchi K, Takahashi S, Kawanami T, Kato T, Sasaki H. [1968] Retinal degeneration in hereditary ceruloplasmin deficiency. *Ophthalmologica* 212: 11–14.
- [102] Patel BN, Dunn RJ, Jeong SY, Zhu Q, Julien JP, David S. [2002] Ceruloplasmin regulates iron levels in the CNS and prevents free radical injury. *J Neurosci* 22: 6578–6586.
- [103] Jeong SY, David S. [2006] Age-related changes in iron homeostasis and cell death in the cerebellum of ceruloplasmin-deficient mice. *J Neurosci* 26: 9810–19.
- [104] Ragan HA, Nacht S, Lee GR, Bishop CR, Cartwright GE. [1969] Effect of ceruloplasmin on plasma iron in copperdeficient swine. *Am J Physiol* 217: 1320–1323.



- [105] Mukhopadhyay CK, Attieh ZK, Fox PL. [1998] Role of ceruloplasmin in cellular iron uptake. *Science* 279: 714–717.
- [106] Attieh ZK, Mukhopadhyay CK, Seshadri V, Tripoulas NA, Fox PL. [1999] Ceruloplasmin ferroxidase activity stimulates cellular iron uptake by a trivalent cation-specific transport mechanism. *J Biol Chem* 274: 1116–1123.
- [107] Xie JX, Tsoi YK, Ke Y, Qian ZM. [2002] Effects of ferroxidase activity and species on ceruloplasmin mediated iron uptake by BT325 cells. *Mol Brain Res* 99: 12–16.
- [108] Ke Y, Ho K, Du J, et al. [2006] Role of soluble ceruloplasmin in iron uptake by midbrain and hippocampus neurons *J Cell Biochem* 98: 912–919.
- [109] Jeong SY, David S. [2003] Glycosyl-phosphatidylinositolanchored ceruloplasmin is required for iron efflux from cells in the central nervous system. *J Biol Chem* 278: 27144– 27148.
- [110] De Domenico I, Ward DM, di Patti MC, et al. [2007] Ferroxidase activity is required for the stability of cell surface ferroportin in cells expressing GPI-ceruloplasmin. *EMBO J* 26: 2823–2831.
- [111] Fox PL, Mazumder B, Ehrenwald E, Mukhopadhyay CK. [2000] Ceruloplasmin and cardiovascular disease. *Free Radic Biol Med* 28: 1735–1744.
- [112] Goldstein IM, Kaplan HB, Edelson HS, Weissmann G. [1979] Ceruloplasmin: A scavenger of superoxide anion radicals. *J Biol Chem* 254: 4040–4045.
- [113] Mukhopadhyay CK, Ehrenwald E, Fox PL. [1996] Ceruloplasmin enhances smooth muscle cell and endothelial cell-mediated low density lipoprotein oxidation by a superoxide-dependent mechanism. *J Biol Chem* 271: 14773– 14778.
- [114] Mukhopadhyay CK, Mazumder B, Lindley PF, Fox PL. [1997] Identification of the prooxidant site of human ceruloplasmin: A model for oxidative damage by copper bound to protein surfaces Proc Natl Acad Sci USA 94: 11546– 11551.
- [115] Shiva S, Wang X, Ringwood LA, et al. [2006] Ceruloplasmin is a NO oxidase and nitrite synthase that determines endocrine NO homeostasis. *Nat Chem Biol* 2: 486–493.
- [116] Daimon M, Yamatani K, Igarashi M, et al. [1995] Fine structure of the human ceruloplasmin gene. *Biochem Biophys Res Commun* 208: 1028–35.
- [117] Hellman NE, Kono S, Miyajima H, Gitlin JD. [2002] Biochemical analysis of a missense mutation in aceruloplasminemia. *J Biol Chem* 277: 1375–80.
- [118] Yang F, Naylor SL, Lum JB, et al. [1986] Characterization, mapping and expression of the human ceruloplasmin gene. *Proc Natl Acad Sci USA* 83: 3257–61.
- [119] Fortna RR, Watson HA, Nyquist SE. [1999] Glycosylphosphatidylinositol- anchored ceruloplasmin is expressed by rat Sertoli cells and is concentrated in detergent insoluble membrane fractions. *Biol Reprod* 61:1042–49.
- [120] Patel BN, Dunn RJ, David S. [2000] Alternative RNA splicing generates a glycosyl-phosphatidylinositol-anchored form of ceruloplasmin in mammalian brain. *J Biol Chem* 275: 4305–10.
- [121] Salzer JL, Lovejoy L, Linder MC, Rosen C. [1998] Ran-2, a glial lineage marker, is a GPI-anchored form of ceruloplasmin. J *Neurosci Res* 54:147–57.
- [122] Banha J, Marques L, Oliveira R, et al. [2008] Ceruloplasmin expression by human peripheral blood lymphocytes: A new

link between immunity and iron metabolism. *Free Radical Biology and Medicine* 44: 483–492.

- [123] Zaitseva I, Zaitsev V, Card G, et al. [1996] The nature of the copper centres in human ceruloplasmin. *J Biol Inorg Chem* 1: 49–63.
- [124] Gitlin JD, Schroeder JJ, Lee-Ambrose LM, Cousins RJ. [1992] Mechanisms of caeruloplasmin biosynthesis in normal and copper-deficient rats. *Biochem J* 282: 835–39.
- [125] Holtzman NA, Gaumnitz BM. [1970] Identification of an apoceruloplasmin like substance in the plasma of copper deficient rats. *J Biol Chem* 245: 2350–53.
- [126] Miyajima H, Adachi J, Kohno S, Takahashi Y, Ueno Y, Naito T. [2001] Increased oxysterols associated with iron accumulation in the brains and visceral organs of acaeruloplasminaemia patients. *QJM* 94: 417–22.
- [127] Kaneko K, Yoshida K, Arima K, et al. [2002] Astrocytic deformity and globular structures are characteristic of the brains of patients with aceruloplasminemia. *Journal of Neuropathology and Experimental Neurology* 61: 1069–77.
- [128] Oide T, Yoshida K, Kaneko K, Ohta M, Arima K. [2006] Iron overload and antioxidative role of perivascular astrocytes in aceruloplasminemia. *Neuropathol Appl Neurobiol* 32: 170–76.
- [129] Kono S, Miyajima H. [2006]. Molecular and pathological basis of aceruloplasmineinia. *Biol Res* 39: 15–23.
- [130] Tapryal N, Mukhopadhyay C, Das D, Fox PL, Mukhopadhyay CK. [2009] Reactive oxygen species regulate ceruloplasmin by a novel mRNA decay mechanism involving its 3'-untranslated region: implications in neurodegenerative diseases. J Biol Chem 284: 1873–1883.
- [131] Wu LJ, Leenders AGM, Cooperman S, et al. [2004] Expression of the iron transporter ferroportin in synaptic vesicles and the blood-brain barrier. *Brain Res* 1001: 108– 117.
- [132] Cairo G, Tacchini L, Pogliaghi G, Anzon E, Tomasi A, Bernelli Zazzera A. [1995] Induction of ferritin synthesis by oxidative stress: transcriptional and post-transcriptional regulation by expansion of the 'free' iron pool. *J Biol Chem* 270: 700–703.
- [133] Mehlhase J, Sandig G, Pantopoulos K, Grune T. [2005] Oxidation-induced ferritin turnover in microglial cells: role of proteasome. *Free Radic Biol Med* 38: 276–285.
- [134] Garneau NL,Wilusz J, Wilusz CJ. [2007] The highways & byways of mRNA decay. *Nat Rev Mol Cell Biol* 8: 113–126.
- [135] He Y, Thong PS, Lee T, et al. [2003] Dopaminergic cell death precedes iron elevation in MPTP-injected monkeys. *Free Radic Biol Med* 35: 540–547.
- [136] Dawson TM, Dawson VL. [2003] Molecular pathways of neurodegeneration in Parkinson's disease. *Science* 302: 819– 822.
- [137] Block ML, Hong JS. [2005] Microglia and inflammationmediated neurodegeneration: multiple triggers with a common mechanism. *Prog Neurobiol* 76: 77–98.
- [138] Lobsiger GS, Cleveland DW. [2007] Glial cells as intrinsic components of non-cell-autonomous neurodegenerative disease. *Nat Neurosci* 10: 1355–1360.
- [139] Kaneko K, Hineno A, Yoshida K and Ikeda SI. [2008] Increased vulnerability to rotenone-induced neurotoxicity in ceruloplasmin-deficient mice. *Neurosci Lett* 446: 56–58.

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